

Kinetic characterization of almond β -glucosidase

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Introduction

β -glucosidase (EC 3.2.1.21) are enzymes which catalyze the hydrolysis of O- β -Glycosidase bonds to terminal non-reducing residues in beta-D-glucosides and oligosaccharides, with release of a molecule of glucose. They are involved in several biological functions [1] in bacteria, fungi, plants and mammals alike. Their capacity to catalyze the inverse reaction of transglycosilation [2] makes them quite interesting as an industrial catalyzer [3,4]. The objective of these experiments was to propose a kinetic mechanism for almond β -glucosidase .

Methods

0,9 ml Substrate pNPG $\xrightarrow[40^\circ\text{C}]{5\text{ min}}$ 0,1ml enzyme $\xrightarrow[40^\circ\text{C}]{\text{time}}$ 1,5 ml NaOH 0,2M

- Standarization of assay conditions

In order to obtain velocities of reaction, we first needed to construct a calibration plot of pNP. After that, assays were run on to determine optimal enzyme concentration and approximated K_m . Then, we checked the linearity of the assay with time.

- Determination of kinetic parameters

Assays were performed in the same conditions previously established and with different substrate conditions. After having obtained initial velocities, macroscopic kinetic parameters were determined.

- Effects of temperature in the catalysis

Kinetic parameters of the enzyme were determined at different temperatures. Activation energy (E_a) was estimated using an Arrhenius plot. Q_{10} coefficient was also calculated. $Q_{10} = V_{max}(T)/V_{max}(T-10)$.

- Inhibition studies

Finally we studied the inhibition of β -glucosidase using different concentrations of glucose (a product of this reaction) and δ - gluconolactone (analogue of the transition state).

Results

- Assay standarization

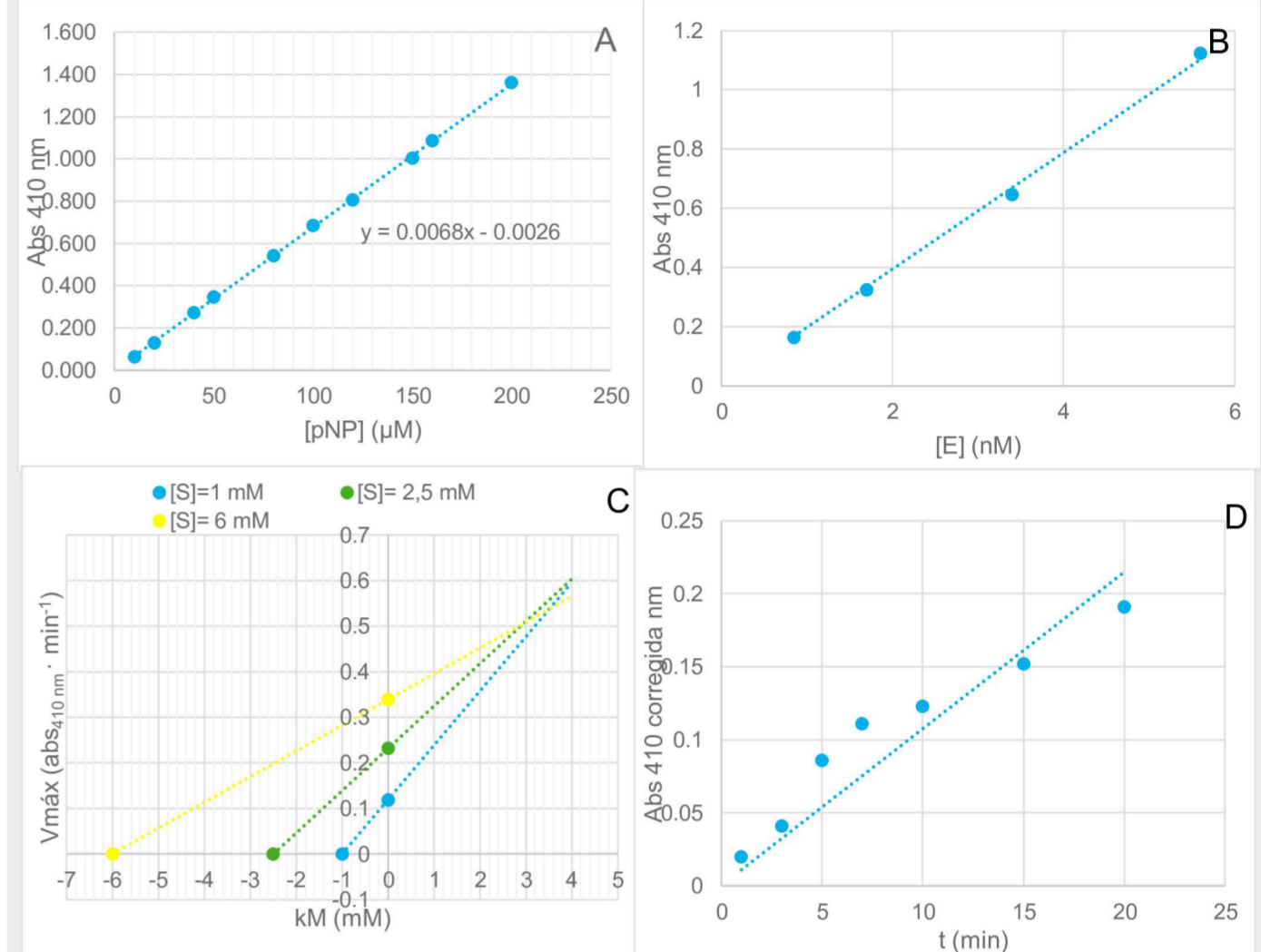


Figure 1. A) pNP calibration plot. B) Optimal $[E]$ (3.5 nM). C) Approximated K_m (3.65 mM). D) Linearity with time

Results

-Kinetic parameters

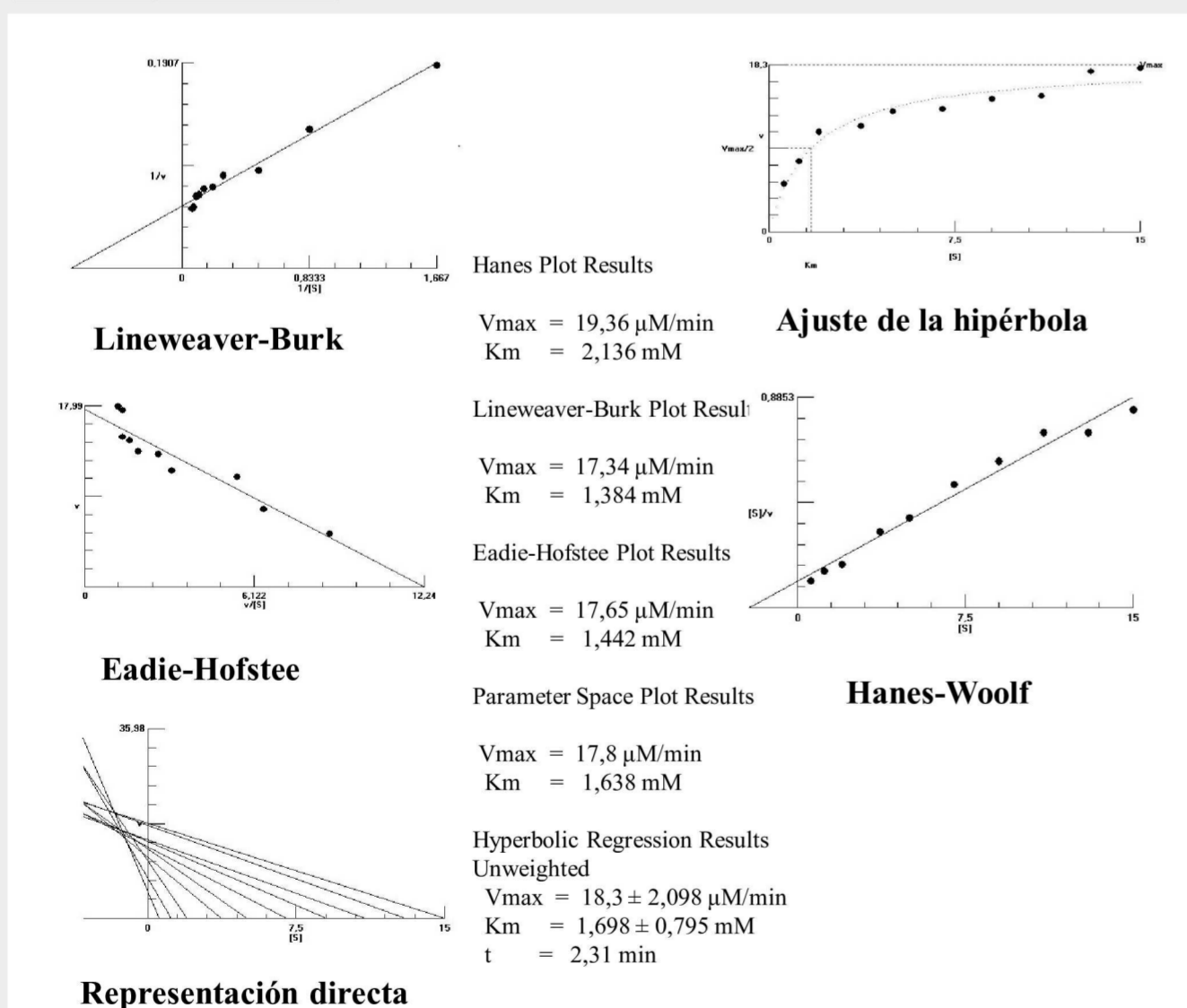


Figure 2. Grafic representations of kinetic parameters and comparative values of them.

-Temperature effect

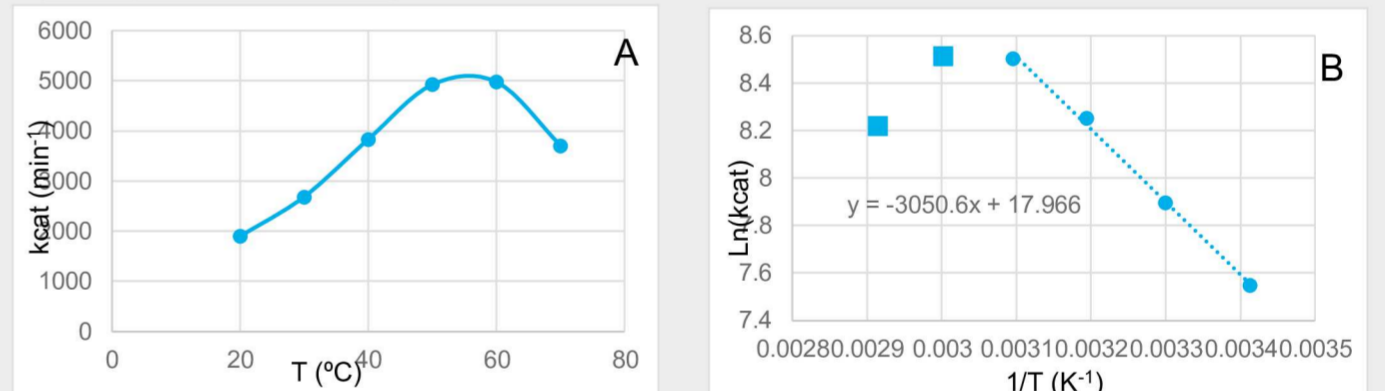


Figure 3. A) Effect of temperature on k_{cat} . B) Arrhenius plot. $E_a = 25.35 \text{ kJ/mol}$. $Q_{10} = 1.43$

-Inhibition studies

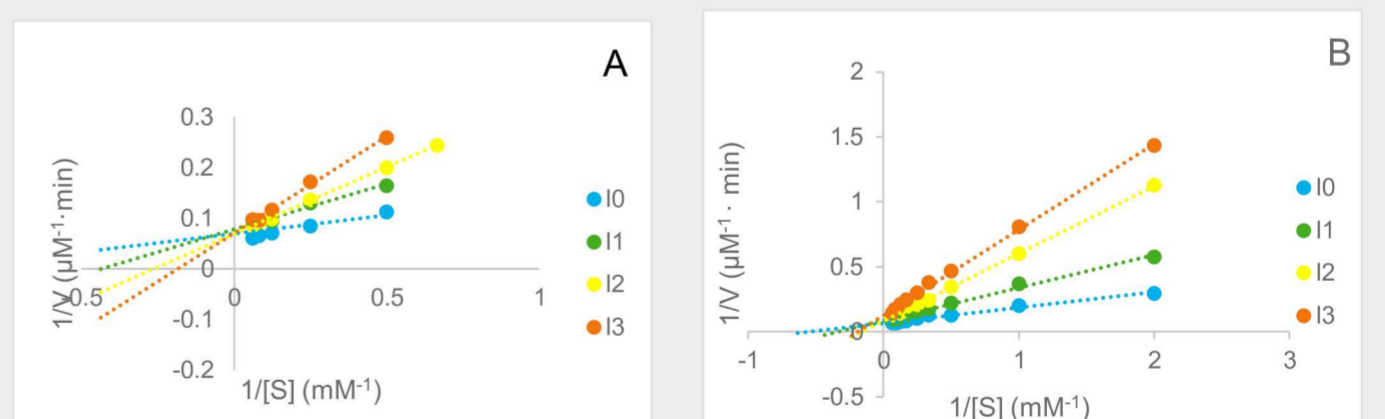


Figure 4. A) Inhibition by δ -gluconolactone ($K_{is} = 0.06 \text{ mM}$). B) Inhibition by glucose ($K_{is} = 205 \text{ mM}$).

Bibliography

- [1]. Bhatia, Y.; Mishra, S.; Bisaria, V. S., Microbial beta-glucosidases: Cloning, properties, and applications. *Critical Reviews in Biotechnology* 2002, 22, 375-407
- [2]. Mladenoska, I.; Grey, C. E.; Winkelhausen, E.; Kuzmanova, S.; Adlercreutz, P., Competition between transglycosylation and hydrolysis in almond beta-glucosidase-catalyzed conversion of p-nitrophenyl-beta-D-glucoside in monophasic water/alcohol mixtures. *Biocatalysis and Biotransformation* 2007, 25, 382-385.
- [3]. Bhat, M. K.; Bhat, S., Cellulose degrading enzymes and their potential industrial applications. *Biotechnology Advances* 1997, 15, 583-620
- [4]. Thuan, N.H.; Sohng, J.K., Recent biotechnological progress in enzymatic synthesis of glycosides. *Journal of Industrial Microbiology and Biotechnology* 2013, 40, 1329-1356
- [5]. Kara, H. E.; Sinan, S.; Turan, Y., Purification of beta-glucosidase from olive (*Olea europaea* L.) fruit tissue with specifically designed hydrophobic interaction chromatography and characterization of the purified enzyme. *Journal of Chromatography B-Analytical Technologies in the Biomedical and Life Sciences* 2011, 879, 1507-1512
- [6]. Seshadri, S.; Akiyama, T.; Opasirij, R.; Kuaprasert, B.; Cairns, J. K., Structural and enzymatic characterization of Os3BGlu6, a rice beta-glucosidase hydrolyzing hydrophobic glycosides and (1-3) and (1-2)-linked disaccharides. *Plant Physiology* 2009, 151, 47-58
- [7]. Itohshida, T.; Hiraiwa, M.; Uda, Y., Purification and properties of beta-D-glucosidase (linamarase) from the butter bean, *Phaseolus lunatus*. *Journal of Biochemistry* 1987, 101, 847-854.

Conclusions

In conclusion, these experiments were held in quite reasonable conditions according to the obtained experimental values for the kinetic parameters, which only differ slightly from other studies [5,6,7], as does the incubation temperature of 40°C . Attending to the inhibition study (a competitive type), and since the ultimate goal of these experiments was to propose a kinetic mechanism for almond β -glucosidase, we concluded this mechanism to be a uni-bi ping-pong type as showed in figure 5.

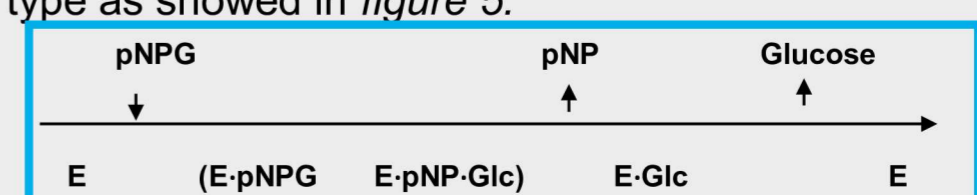


Figure 5. Cleland scheme